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## Revised structure of deacetyl-1,10-didehydrosalvinorin G

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**Abstract**—In comparison with the NMR data of salvinorin A and its 8-epimer, the published structure of deacetyl-1,10-didehydrosalvinorin G was revised to its 8-epimer. The stereochemistry of 8-*epi*-deacetyl-1,10-didehydrosalvinorin G was further confirmed by NOESY and chemical synthesis.

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Salvinorin A (1a), a non-nitrogenous neoclerodane diterpenoid, was isolated from the Mexican medicinal plant *Salvia divinorum*.<sup>1,2</sup> Compound 1a was identified as a potent and selective kappa ( $\kappa$ ) opioid receptor (KOR) agonist and as the key ingredient for psychoactive effects.<sup>3–5</sup> During the course of structure–activity relationship (SAR) studies,<sup>6–18</sup> it was found that 1a and its derivatives readily underwent epimerization at C-8 under basic conditions. Using various inorganic bases, such as NaBH<sub>4</sub>,<sup>2,12,19</sup> NaHCO<sub>3</sub>,<sup>20</sup> Na<sub>2</sub>CO<sub>3</sub>,<sup>7,12</sup> K<sub>2</sub>CO<sub>3</sub>,<sup>8</sup> LiSEt<sup>10</sup> and LiI,<sup>11</sup> 1a produced corresponding natural (8-H $\beta$ ) and unnatural (8-H $\alpha$ , also called 8-*epi*-) mixtures. Surprisingly, treatment of 1a with excess strong base KOH or Ba(OH)<sub>2</sub> gave a natural salvinorin derivative deacetyl-1,10-didehydrosalvinorin G that was claimed by different research groups to have structures **2a** and **3**.<sup>13,8</sup> In general, the affinity and potency of natural salvinorin derivatives show much higher KOR binding activities than those of unnatural 8-epimers.<sup>8,12,15,21</sup> Since the configuration of a molecule can significantly affect KOR binding, it is essential to establish a reliable method which would unambiguously determine the stereochemistry at C-8 of salvinorins. This prompted us to conduct a comprehensive analysis of the <sup>1</sup>H and <sup>13</sup>C NMR data of natural and unnatural salvinorin derivatives. The differences of **1a** and its 8epimer (**1b**) in their <sup>1</sup>H and <sup>13</sup>C NMR spectra are discussed in this Letter. Based on the summarized NMR data and chemical synthesis, the stereochemistry of **2a** at C-8 is revised to its 8-epimer (**2b**) as shown in Figure 1.



Figure 1. Conformational structures of 2a and 2b.

Keywords: Salvinorin A; Epimer; NMR; Revised structure.

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In connection with our previous study related to determining the stereochemistry of betulinic acids,<sup>22</sup> we reco-gnized that the <sup>1</sup>H and <sup>13</sup>C NMR data of salvinorins and their corresponding 8-epimers have significant differences. A pair of 8-epimers (1a and 1b), isolated<sup>9</sup> and synthesized<sup>8</sup> in our group, was selected as standard compounds for NMR analyses. Using 2D NMR techniques, including COSY, NOESY, HMQC and HMBC, permitted the full assignments of all <sup>1</sup>H and <sup>13</sup>C NMR chemical shifts. The relative stereochemistry at the C-8 position of these compounds was unambiguously determined, based on their <sup>1</sup>H and <sup>13</sup>C NMR data. For instance, in the <sup>1</sup>H NMR spectra, the C-8-H of 1a resonated at  $\delta$  2.07 (dd, J = 3.0 and 12.0 Hz), confirming axial orientation, while that of **1b** resonated at  $\delta$  2.45 (d, J = 2.7 Hz) for equatorial orientation. The chemical shift change ( $\Delta\delta$ ) between **1a** and **1b** is about 0.38 ppm (Table 1). In addition, the C-12-H of 1b shifted upfield  $(\Delta \delta \ 0.27 \text{ ppm})$  to  $\delta \ 5.26$  (dd, J = 1.8 and 12.0 Hz) in comparison with that of **1a** at  $\delta$  5.53 (dd, J = 4.8 and 11.7 Hz). The coupling constants of H-12 revealed the axial orientation of H-12 in both 1a and 1b. The

Table 1.  $^{1}\mathrm{H}$  NMR data (300 MHz) at H-8 and -12 for 1a, 1b and 2b in CDCl\_3

Compound	1a <sup>a</sup>	1b <sup>a</sup>	$\Delta \delta_{1b-1a}$	2b
Η-8 (δ)	2.07	2.45	+0.38	2.99
J value	dd, 3.0, 12.0	d, 2.7		dd, 5.1, 9.6
H-12 $(\delta)$	5.53	5.26	-0.27	5.45
J value	dd, 4.8, 11.7	dd, 1.8, 12.0		dd, 2.4, 12.3

<sup>a</sup> See Refs. 10, 20 and 23 for full <sup>1</sup>H NMR assignments of **1a** and **1b**.

Table 2. <sup>13</sup>C NMR data (75 MHz) for 1a, 1b and 2b in CDCl<sub>3</sub>

X-ray analyses showed that the lactone ring in 1a adopts a chair confirmation, 1,2,17 while the one in **1b** is a boat conformation.<sup>21</sup> The different conformation may explain the J values differences of H-12 between 1a and 1b. On the other hand, the H-8 configuration also strongly affects the <sup>13</sup>C NMR data in B- and C-ring carbons. For instance, the methylene carbon (C-11), carbonyl carbon (C-17) and axial methyl carbon (C-20) of 1b showed lower-field chemical shifts in comparison with those of **1a** (Table 2), while the <sup>13</sup>C resonances of C-6, C-8, C-12, C-13 and C-19 of 1b shifted to upper field. In summary, the characteristic carbon peaks of C-12, C-17 and C-20 can be employed readily for identification of natural and unnatural 8-epimers. In the previous published papers,<sup>10,13,21</sup> 8-epimers were mainly determined based on the coupling constants of H-8 and H-12, irradiation of H-12 for a NOE enhancement of H-8, and a general TLC Rf value. Our generalized NMR data should provide reliable information for identification of future salvinorin analogs, including C-8 epimers.

We reported previously that treatment of 1a with Ba(OH)<sub>2</sub> in MeOH gave an unexpected oxidative-elimination product that was assigned as structure  $3.^8$  In a very short time span, the structure of 3 was revised to deacetyl-1,10-didehydrosalvinorin G (2a), which was synthesized in 1 M KOH methanol solution.<sup>13</sup> The structural revision was based on extensive NMR experiments, including <sup>1</sup>H NMR, <sup>13</sup>C NMR, DEPT, COSY, HMQC, HMBC and NOE, and HR-ESI-MS. Following the published procedure (Scheme 1),<sup>13</sup> indeed, we isolated the same product.<sup>24</sup> The NMR data were in full

Carbon #	<b>1</b> a	1b	$\Delta \delta_{\mathbf{1b}-\mathbf{1a}}{}^{\mathrm{a}}$	2b	Carbon #	<b>1</b> a	1b	$\Delta \delta_{\mathbf{1b}-\mathbf{1a}}^{\mathbf{a}}$	2b
1	202.0	202.3		145.1	13	125.2	123.4	-1.8	124.4
2	75.0	75.2		180.7	14	108.4	108.5		108.4
3	30.7	30.7		128.2	15	143.7	143.5		143.7
4	53.6	52.9		157.5	16	139.4	139.7		139.6
5	42.1	42.2		42.3	17	171.1	173.4	+2.3	173.2
6	38.1	34.0	-4.1	28.3	18	171.5	171.8		165.4
7	18.1	17.6		21.9	19	16.4	15.2	-1.2	30.3
8	51.4	45.2	-6.2	44.8	20	15.2	24.6	+9.4	24.4
9	35.4	34.7		37.6	-COOMe	52.0	51.7		52.6
10	64.1	64.0		140.0	$-COCH_3$	170.0	169.7		
11	43.4	48.0	+4.6	36.8	$-COCH_3$	20.6	20.5		_
12	72.0	70.0	-2.0	70.8					

<sup>a</sup> Data is given when  $\Delta \delta_{1b-1a}$  is more than 1.0 ppm.





Figure 2. Key NOE interactions of 2b.

agreement with previous report<sup>13</sup> except that the assignments of H-7 $\alpha$  ( $\delta$  1.98) and H-7 $\beta$  ( $\delta$  2.24) should be reversed as H-7 $\alpha$  ( $\delta$  2.24) and H-7 $\beta$  ( $\delta$  1.98). After careful comparison of our <sup>1</sup>H and <sup>13</sup>C NMR data with those of 1a and 1b (Tables 1 and 2), we found that structure 2a assigned to the product was incorrect,<sup>13</sup> and the correct structure should be 8-epi-deacetyl-1,10-didehydrosalvinorin G (2b, Fig. 1). In the <sup>1</sup>H NMR spectrum, the C-8-H of **2b** shifted much lower field to  $\delta$  2.99 (dd, J = 5.1 and 9.6 Hz), while the C-12-H shifted upfield slightly compared with that of 1a (Table 1). The H-12 coupling constants (dd, J = 2.4 and 12.3 Hz) of **2b** were closer to those of 1b (dd, J = 1.8 and 12.0 Hz) than of 1a (dd, J = 4.8 and 11.7 Hz), but the H-8 of **2b** showed the J values (5.1 and 9.6 Hz) for axial orientation.<sup>13</sup> However, this perplexing configuration at C-8-H was soon resolved by comparing the C-ring <sup>13</sup>C NMR data of 2b with those of 1a and 1b (Table 2). The <sup>13</sup>C NMR chemical shifts at C-8, C-12, C-17 and C-20 are very similar to those of 1b, indicating that the orientation of H-8 is  $\alpha$ . The revised structure 2b was further confirmed by the NOE interactions shown in Figure 2. In the NOESY spectrum (see Supplementary data), H-8 ( $\delta$  2.99) showed cross peaks to H-7 $\alpha$  ( $\delta$  2.24, strong), H-7 $\beta$  ( $\delta$  1.98, weak), H-19 ( $\delta$  1.72) and H-20 ( $\delta$  1.67), while H-12 ( $\delta$  5.45) related to H-11 $\alpha$  ( $\delta$  3.11, strong), H-11 $\beta$  ( $\delta$  2.02, weak) and H-20 ( $\delta$  1.67). It should be noted that the crossed peak between H-8 and H-12 was very weak. Furthermore, when **1b**, the 8-epimer of **1a**, was treated with 1 M KOH in MeOH, it afforded endione **2b** in a 67% isolated yield (Scheme 1).<sup>24</sup> Valdes et al.<sup>2</sup> reduced **1a** with NaBH<sub>4</sub> in *i*-PrOH and afforded equal amounts of 8-epimeric mixtures (diol **4a** and 8*epi*-diol **4b**), while Munro et al.<sup>13</sup> reduced **2a** with NaBH<sub>4</sub> in EtOH/CH<sub>2</sub>Cl<sub>2</sub> and obtained sole product— 8-*epi*-diol **4b**. All of this evidence supports structure **2b**.

The coupling constants of H-8 in 2b were misleading in comparison with those of other 8-epi-salvinorins. This is the main reason for Munro et al.<sup>13</sup> to determine H-8 as  $\beta$ configuration. Because of the double bond between C-1 and C-10, the conformation of 2b is distorted. It is known that A/B/C rings in 1a are in chair/chair/chair conformation<sup>1,2</sup> and A/B/C rings in **1b** are in chair/ chair/boat conformation.<sup>21</sup> Based on NOESY and molecular modeling analyses, A/B/C rings in 2b should be face-down boat/twist-chair/twist-chair conformation (Fig. 1). Under this circumstance, both H-7ß and H-8 resemble axial orientations and showed a closer diaxial coupling constant (9.6 Hz). Without sufficient NOE data,<sup>13</sup> the incorrect assignments of H-7 $\alpha$  and H-7 $\beta$  in **2b** might depend on those of the known salvinorins C (5a), D (5b), E (5c) and F (5d), isolated from S. divino-rum by Munro and Rizzacasa.<sup>25</sup> Because the chemical shifts of H-7 and H-8 in 5a, 5b, 5c and 5d overlapped,<sup>19,25</sup> the unambiguous assignments of H-7 $\alpha$  and H-7β with NOESY spectra became very difficult. Unexpectedly, one year later, Munro gave the revised assignments of H-7 $\alpha$  and H-7 $\beta$  of **2b** without any scientific explanation.<sup>20</sup> Based on our NOE data, all protons in **2b** were fully assigned.<sup>24</sup>

In conclusion, a concise and informative NMR method for the determination of the C-8-H configuration of salvinorins was established. Based on this method, the correct product obtained by the treatment of **1a** with hydroxide in MeOH has been identified. The C-8



epimeric structure (**2b**) was confirmed by NOESY spectrum and chemical conversion.

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## Supplementary data

Copies of <sup>1</sup>H and <sup>13</sup>C NMR spectra of **1a**, **1b** and **2b**, and 2D NOESY spectrum of **2b**. Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tetlet.2007.05.179.

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- 24. Synthesis of **2b**. Compound **1a** (40 mg, 92.6 µmol) was added to a solution of KOH in MeOH (1 M, 3 mL) and stirred at room temperature for 1 h. Following the previous procedure,<sup>13</sup> crude **2b** was obtained. Further purification by silica gel column [hexane/AcOEt (2:1)] gave pure **2b** (12 mg, yield 34%). In the same manner, **2b** was also obtained from **1b** in 67% isolated yield. The <sup>1</sup>H and <sup>13</sup>C NMR chemical shifts of **2b** were in full agreement with those of published data.<sup>8,13</sup> The H-6 $\alpha$  and H-6 $\beta$  were assigned as  $\delta$  2.53 and  $\delta$  1.67–1.77, respectively. The H-7 $\alpha$  and H-7 $\beta$  were revised to  $\delta$  2.24 and  $\delta$  1.98, respectively. EI-MS m/z: 386 (M<sup>+</sup>).
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